Stealth Adaptation of Viruses: Implications for Therapy and for Potential Toxicity of Vaccines

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Abstract

The scope of human and animal virology research is likely to undergo a major expansion. It will occur with the realization that the immune system may fail to effectively recognize derivative viruses, which no longer express the major antigens normally targeted by cytotoxic T lymphocytes (CTL). This immune evasion mechanism has been termed stealth adaptation. Persistent infections with stealth adapted viruses can lead to various illnesses and especially those with neuropsychiatric symptoms. Certain stealth adapted virus infections almost certainly entered into humans via monkey cytomegalovirus contaminated polio vaccines. Preexisting stealth adapted virus infections may render individuals susceptible to disease provocation from administered vaccines. On a more promising note, the research has helped identify an alternative cellular energy (ACE) pathway that can suppress both stealth adapted and conventional viruses. The ACE pathway is providing a new therapeutic paradigm that extends beyond virology to general medicine. It is also leading towards a basic understanding of how Nature prevents the fusion of opposite electrical charges. It is postulated that Nature does so through a fundamental force, tentatively termed KELEA (kinetic energy limiting electrostatic attraction).

Keywords: ACE, alternative cellular energy, African green monkey simian cytomegalovirus, AIDS, autism, brain, cytomegalovirus, inflammation KELEA, PCR, polymerase chain reaction, polio vaccine, SCMV, stealth adapted viruses


Introduction

Inflammation is generally regarded as an essential component of an effective defense mechanism against virus infections [1]. While it can be directly evoked by virus induced cellular damage, inflammation is mainly viewed as an outcome of cellular immunity [2] and, in particular, T lymphocyte recognition of virus coded components (antigens). Since each individual T lymphocyte is genetically programmed to recognize only a single antigenic specificity [3], effective lymphocyte engagement with a virus infected cell requires multiple copies of the recognized antigen to be displayed on the surface of the infected cells. There are additional requirements for a virus component to be recognized by T lymphocytes, including the need to bind to a particular region of the major histocompatibility complex (MHC) of the infected cell [4]. These restrictions account for the finding that relatively few virus components actually comprise major targets for the cellular immune response. Among the best examples is human cytomegalovirus (HCMV) in which one component (coded by the UL83 gene) is targeted by over 50% of anti-HCMV cytotoxic T lymphocytes (CTL) [5]. The majority of the remaining responding CTL are directed against either the UL55 or UL123 gene coded antigen [6]. Another example of immune evasion is with hepatitis B virus (HBV), in which a mutation can occur in the gene coding the CTL targeted hepatitis surface antigen [7]. This example is not as clear as with HCMV, however, since the nucleotides coding the surface antigen also code the DNA polymerase gene, often rendering the mutant virus less virulent and less sensitive to anti-viral therapy [7]. A generic immune evasion mechanism that results from the loss or mutation of components normally targeted by CTL has been termed stealth adaptation [8-11]. A lesser form of this mechanism can be antigenic loss or mutation that primarily limits immunological recognition in those individuals with certain MHC specificities.

The existence of stealth adapted viruses has shed important new light onto many important aspects of human and animal health. This review i) provides a brief introduction to the early identification of stealth adapted viruses, including evidence that some are derived from African green monkey simian cyto
confirmed abundant virus particles, along with extensive vacuoles transmitted to secondary cultures [17]. Electron microscopy patient. After several weeks, fibroblasts inoculated with cells effort was made to culture the blood of the strongly positive PCR disabled with a diagnosis of CFS. While prior viral cultures on encephalitis. The woman subsequently became chronically 43-year-old woman recently hospitalized to rule out meningitis/hemorrhages, hepatomegaly and thrombocytopenia. He was 30% of CFS patients. Reactivity was also seen in the cerebrospinal fluid (CSF) repeatedly obtained from an infant with choroid hemorrhages, hepatomegaly and thrombocytopenia. He was suspected of having a viral illness at birth but routine virus cultures were negative. A positive PCR was also obtained on a 1990 stereotactic brain biopsy of a school teacher with declining cognition and whose MRI showed bilateral periventricular lesions [17]. In spite of the positive PCR, the brain biopsy was totally devoid of any inflammation, although cellular damage was apparent histologically and confirmed by electron microscopy. Even though intact virus particles were not seen, there was extensive vacuolization, mitochondrial disruption and unusual intracellular inclusions, consistent with virus pathology [17]. Because of the lack of any discernable inflammation, the putative virus being detected by the PCR was designated as being a stealth virus. The same designation was used to explain positive PCR results on several acellular CSF samples obtained from patients with severe, chronic neurological illnesses and on blood samples from CFS patients.

Stealth Adapted African Green Monkey Simian Cytomegalovirus (SCMV)

Evidence for active virus infections in CFS patients was initially obtained using the polymerase chain reaction (PCR) as a diagnostic tool [16]. In very early (pre-1990) studies, low stringency PCR using primers designed to broadly detect herpes viruses yielded weak but still positive findings in approximately 30% of CFS patients. Reactivity was also seen in the cerebrospinal fluid (CSF) repeatedly obtained from an infant with choroid hemorrhages, hepatomegaly and thrombocytopenia. He was suspected of having a viral illness at birth but routine virus cultures were negative. A positive PCR was also obtained on a 1990 stereotactic brain biopsy of a school teacher with declining cognition and whose MRI showed bilateral periventricular lesions [17]. In spite of the positive PCR, the brain biopsy was totally devoid of any inflammation, although cellular damage was apparent histologically and confirmed by electron microscopy. Even though intact virus particles were not seen, there was extensive vacuolization, mitochondrial disruption and unusual intracellular inclusions, consistent with virus pathology [17]. Because of the lack of any discernable inflammation, the putative virus being detected by the PCR was designated as being a stealth virus. The same designation was used to explain positive PCR results on several acellular CSF samples obtained from patients with severe, chronic neurological illnesses and on blood samples from CFS patients.

Through serendipity, an additional primer set originally matched to a retrovirus was also tested in some of the patients. It yielded a strikingly strong PCR using blood samples from a 43-year-old woman recently hospitalized to rule out meningitis/encephalitis. The woman subsequently became chronically disabled with a diagnosis of CFS. While prior viral cultures on CFS patients were providing suggestive results, a determined effort was made to culture the blood of the strongly positive PCR patient. After several weeks, fibroblasts inoculated with cells from her blood displayed an overt cytopathic effect (CPE); easily transmitted to secondary cultures [17]. Electron microscopy confirmed abundant virus particles, along with extensive vacuoles and inclusions [18]. A similar very strongly positive culture was obtained from the CSF of a 21-year-old female patient admitted in coma to the Los Angeles County Hospital. The patient had a 4-year history of a severe manic-depressive (bipolar) psychosis [19]. Remarkably, her CSF was essentially acellular and her coma was clinically attributed to anoxia from a drug overdose. Aliquots of the positive culture were sent to the Los Angeles County Public Health Laboratory. They simply confirmed the CPE and lack of staining by available virus typing antisera. Minor changes in the culturing procedures led to more rapid and consistently positive cultures in CFS patients referred by Dr. Jay Goldstein, a physician specializing in CFS patients. The culture CPE was characterized by the formation of foamy, vacuolated cells with synctyia [18]. The finding of positive cultures in CFS patients was reported to the Los Angeles County Public Health Laboratory.

PCR generated products from the strongly positive culture of the 43-year-old CFS patient were cloned and sequenced. The sequencing identified two distinct nucleotide sequences, each flanked at both ends by the same primer [18]. The sequences of one of the clones showed highly significant matching, but not complete homology, to HCMV sequences available on GenBank [18]. At the time, GenBank contained no significant matching sequences to the other clone. DNA extracted from pelleted culture supernatant, with and without subsequent agarose banding, was also cloned into plasmids. Sequencing allowed for the design of additional PCR primers that yielded strongly positive reactions when tested on repeated cultures from the patient and also on the CSF and on the CSF culture of the patient with the bipolar illness [19]. Other patients’ strongly positive cultures tested negative using the same sets of primers, indicating that different viruses could undoubtedly undergo stealth adaptation.

Sequencing of additional clones from the culture DNA allowed for more comparisons with the available data on GenBank. Near perfect matching was obtained between some of the cloned DNA and the DNA of African green monkey simian cytomegalovirus (SCMV) [20]. Since African green monkeys were used in the manufacturing of polio vaccines, the results were reported to the Food and Drug administration (FDA), Centers for Disease Control and Prevention (CDC) and the polio vaccine manufacturer.

Finer sequencing of the DNA clones showed considerable genetic instability, with mutations, deletions and recombinations [21]. Evidence for the genes corresponding to UL55 and UL83 in HCMV were not found, although the cloning may have failed to capture the UL83 gene because of a paucity of restriction sites of the enzymes used in the cloning. Major mutations were noted in the sequenced region to the SCMV gene corresponding to UL123 in HCMV [9]. The virus genome was also apparently fragmented, with aggregate sequences far beyond the approximate 20 kilobase size of the extracted DNA on agarose [18]. Other data raised the possibility of DNA fragments being bridged by RNA sequences. Moreover, additional sequences were identified in the cloned DNA of purified virus culture supernatant. Some of these sequences matched to cellular DNA [22], while others matched to bacteria [23]. The latter observation is particularly important since it may explain some of the atypical bacteria isolated from culture positive patients [24].

The cultured virus is pathogenic when inoculated into cats producing an acute, severe generalized disease, with marked neurobehavioral disturbance [25]. Yet, there was no
inflammation, justifying the notion of the virus being stealth. Yet from about 6 weeks to 16 weeks when the last cat was necropsied, the remaining cats showed a rather remarkable clinical recovery.

Cytomegalovirus Contamination of Polio Vaccines

The prospect of developing a polio vaccine was given an enormous boost when John Enders and co-workers showed that the polio virus could be successfully cultured [26]. Their tissue culture efforts were facilitated by the earlier discovery of antibiotics. An initial concern in the development of polio vaccines was that any established cell line might be perceived as having undergone genetic changes towards becoming cancerous. In spite of publicly aired advice to the contrary, the decision was, therefore, made to use primary cultures of monkey kidney cells. Rhesus monkeys were initially used by the three early pioneers, Drs. Jonas Salk, Albert Sabin and Hilary Koprowski [27]. Dr. Salk opted to use formalin inactivated viruses as had been used in influenza vaccine development. Because the virus was to be inactivated, he chose to use the virulent disease causing virus strains. The other two researchers opted to use attenuated (less virulent) strains, as modeled after the yellow fever vaccine. Dr. Sabin followed the suggestion of Dr. Roberto Dulbecco and plaque-purified his isolates, gaining an advantage over the work of Dr. Koprowski. Dr. Salk was given the upper hand in the United States, while Dr. Sabin focused his work in the Soviet Union. Dr. Koprowski did studies in both Europe and Africa [27].

The United States preference for Salk vaccine was consistent with concerns expressed by several prominent virologists that using cells directly cultured from monkeys to produce non-inactivated polio viruses posed risks for contamination with monkey viruses. In their competition, Dr. Sabin both published and personally communicated with Dr. Koprowski that the CHAT vaccine produced in rhesus monkeys was contaminated with a difficult to culture cytopathic virus [28]. The CHAT vaccine was subsequently shown to have DNA of rhesus cytomegalovirus (RhCMV) [29]. The CHAT vaccine was extensively tested in chimpanzees, many of which became sick, as did some of the African animal handlers. A plausible case has been made that HIV arose from some of the CHAT vaccine inoculated chimpanzees [29].

A setback to Dr. Salk’s vaccine was that the virus inactivation process was not always successful and polio outbreaks began to occur during springtime, which some attributed to the experimental use of the vaccine. This issue became more transparent with the licensure of the Salk vaccine when many children became infected with an inadequately inactivated vaccine. A bigger setback was the reluctant acceptance that even with standard inactivation, the Salk vaccine still contained a polyoma-like virus. It was identified as simian virus-40 (SV-40), prevalent in rhesus monkeys [27].

Two major changes then ensued in the early 1960’s. The first was to switch from using rhesus monkeys to the use of African green monkeys. The second was to adopt Dr. Sabin’s live, attenuated polio strains [27]. The US Government was challenged by a pharmaceutical company that if they were to license Dr. Sabin’s vaccine, they would have to license Dr. Koprowski’s vaccine. Even though it was being used overseas, the US Government was aware of the questionable virulence and additional contamination issues of Dr. Koprowski’s vaccines. The Government appealed to Dr. Sabin and agreed that he would freely provide his vaccine strains to Lederle as a means of avoiding the forced US licensure of Dr. Koprowski’s vaccine.

The rather abrupt switch from rhesus to African green monkeys and from inactivated to live virus was a continuing cause of concern, even to workers within FDA’s Bureau of Biologics. An employee’s suggestion that sera from the monkeys be tested for antibody reactivity on the cultured cells as a marker for likely virus contamination was opposed by those in authority. Still in 1972, an earnest effort was made to more closely examine monkey kidney cell cultures. Rather than being used for vaccine production, kidney cell cultures from eleven monkeys were examined for contaminating viruses. All eleven cultures produced SCMV, with only 4 of the positive cultures being detectable using the routine screening assay. The vaccine manufacturer formulated contingency plans to respond to any actions of the FDA to the findings of their joint study. A major argument was that extensive experience with the vaccine had not shown signs of any untoward acute infections. Another argument was that contaminating SCMV would be destroyed during passage through the stomach. Possibly the most telling comment was the expressed opinion that FDA would not have the courage to take their product off the market.

Disregarding the emerging evidence for delayed acting (slow) virus-like illnesses, FDA resolved the issue by suggesting if the vaccine cultures were harvested quickly enough and cultures inoculated with anti-polio neutralized vaccine were examined early enough, evidence for SCMV could be avoided. With the later publication of SCMV DNA still being present in vaccines produced after 1972 [29,31], this approach was clearly ineffective.

One of the FDA officials involved in the earlier study was informed in 1978 of foreign (non-monkey) DNA being present in polio vaccine lot. The information was summarily dismissed by the FDA official with the comment “every time you eat an apple you consume foreign DNA”. The particular vaccine lot (3-444) was selected because polio virus neutralized vaccine induced an atypical CPE and showed evidence of reverse transcriptase, initially suggestive of a possible retrovirus. Further studies helped dismiss the likelihood of a retrovirus and the vaccine lot was approved.

Reports of stealth adapted SCMV contributed to efforts to switch back to using formalin inactivated vaccines. There was also the more politically accepted argument that the live, attenuated vaccine can very occasionally revert to a more virulent strain [32]. Still, the problem is that once a virus is introduced into humans, it can persist due to human to human transmission, and in the case of stealth adapted viruses, reciprocal human to animal transmissions. A very major concerned with down-sized stealth adapted viruses is the possibility of virus carriage through bacteria.
Among the body’s organs, the brain is unique in the spatial non-uniformity of its many functions and the cooperative networking between different regions. Even limited localized damage to the brain can, therefore, be far more functionally disruptive than direct virus damage to other organs. While stealth adapted virus infection is widespread in the body, as in the inoculated cats, it is not surprising, therefore, that the predominant manifestations are neurobehavioral. During several years of cultivating for stealth adapted viruses, it was truly exceptional to obtain negative cultures in patients with severe neurological and/or psychiatric illnesses. While typically not as striking and certainly not as frequent, positive results were occasionally found in apparently symptom-free individuals. In several blinded studies, about 10% of control cultures were identified as being clearly positive. A similar incidence of positive cultures was obtained in a study performed in 2002 on blood donors at the University of California Irvine. The implication of this finding in terms of safety of the Nation’s blood supply led the CDC to sponsor efforts to limit further clinical testing.

Many viruses can cause illnesses in only a subset of infected patients. With polio virus, it is generally estimated that only one percent of those infected develop neurological symptoms [27]. If a cytopathic virus is isolated from the blood and particularly from the CSF of a patient with a neuropsychiatric illness, the virus should be considered a likely contributing factor to the patient’s illness until proven otherwise.

A useful insight emerged from the virus studies. It was that PCR and immune typing assays on different cultures yield variable results consistent with stealth adaptation being a generic process, potentially occurring with all human and animal viruses. The heterogeneity of the origins of stealth adapted viruses, along with evidence of genetic deletions and mutations, can render PCR and immune typing assays somewhat unreliable when performed on patients’ blood samples. A negative PCR assay directed to a limited region within a virus type, does not necessarily exclude the possibility that other regions of that type of virus are present or that another type of virus is infecting the patient. Some stealth adapted DNA viruses may revert to replicating RNA forms such that a reverse transcription step needs to be included in the PCR assay [33]. Genetic instability of stealth adapted viruses [21] can also yield antigens with a broader range of immune reactivity than seen with the particular type of virus from which the stealth adapted virus is derived. Mycoplasma and alphaproteobacteria sequences were identified in cultures of the prototype SCMV-derived stealth adapted virus [10]. The likelihood of bacterial sequences in other stealth adapted viruses may explain misleading positive assays for Borrelia burgdorferi, (the cause of Lyme disease) and/or for Mycoplasma fermentans, as variously reported in CFS patients [34,35]. For these reasons, the most reliable detection method for stealth adapted viruses still remains the culturing of blood, CSF and tissue samples from patients and observing for a characteristic CPE, which tends to repair with the production of extracellular materials. The prototype stealth adapted virus grows readily in cell lines from multiple species and has even been grown on an insect cell line. This is consistent with interspecies transfer of infection between patients and their domestic pets [25].

While the majority of patients on whom blood testing was performed were diagnosed as having CFS, autism or a major neuropsychiatric illness [17, 36-38], additional studies involved other patient groups. For example, positive cultures were regularly seen in patients with multiple myeloma [38]. Upon review, many multiple myeloma patients have prior or even ongoing neurocognitive impairments, not typically addressed by their hematologist [39]. Hodgkin’s disease, aggressive lymphomas, glioblastoma, breast and salivary gland tumor patients [40] have also provided examples of strikingly positive stealth adapted virus cultures. Again upon clinical review, mental fatigue and other indicators of brain dysfunction, not uncommonly preceded the patient’s cancer diagnosis. Post chemotherapy fatigue is explainable as an adverse effect of therapy, but breast cancer patients commonly report persisting fatigue even after the cancer has been excised and no chemotherapy administered (personal communication from a patient support group).

A social contact of the female patient from whom the prototype SCMV-stealth adapted virus was isolated, was experiencing many of the same neurocognitive difficulties as the patient. Their viral cultures were essentially identical in appearance. While the female patient was HIV negative, her friend was HIV positive and died shortly thereafter. Positive stealth adapted virus cultures have also been seen in other HIV infected patients and may contribute to the pathogenicity of HIV [41] and particularly to the neurocognitive impairments seen in many AIDS patients [42].

Evidence of non-sexual transmission of stealth adapted viruses has also been observed in community, workplace, family and personal interactions. A significant epidemic, reported to Public Health authorities, occurred in 1996 in Needles, California and in the adjacent Mohave Valley [43]. The epidemic may have originated from returning veterans since several were included among the earlier identified patient population. Included in the epidemic was an infected child who was initially diagnosed as having a behavioral disorder. A detailed neurological examination was conducted after he informed his physician mother of temporarily seeing double (diplopia) [44]. A biopsy of his brain was obtained on the basis of periventricular lesions being seen on MRI. The biopsy showed several foamy vacuolated cells, but without accompanying inflammation [44]. A second brain biopsy was also examined by electron microscopy. It confirmed marked mitochondrial disruption and structured intracellular inclusions, again in the absence of any inflammation [45]. His virus cultures were repeatedly positive, as were his mother’s. Although he responded initially to ganciclovir, he subsequently died within a year of having the first brain biopsy. The mother, a Gulf War veteran, has been persistently ill from the time of the epidemic.

Another virus culture patient believed he acquired an illness from non-intimate socializing during a business trip. He was initially concerned that he had contracted HIV but tested negative. He became more concerned when he realized his son was not behaving normally and that after a weekend visit, his
father became ill. He surreptitiously had his blood and that of his father tested for SCMV by PCR at a primate virology laboratory. Both samples were reported as being positive for SCMV. He also tested positive in culturing for stealth adapted viruses. An employee at the CDC was informed of the results and arranged for a blood sample to be sent to an outside laboratory, which presumably failed to obtain positive results.

Based on positive virus cultures of many children with autism [46] and on some of the mothers, it is confidently postulated that transplacental and possibly postnatal transmission of stealth adapted viruses can lead to the impaired limitations in social interactions, including language skills that characterizes autism. Stealth adapted virus infection can also explain the relatively high frequency of epilepsy seen in children with autism. The severity and actual manifestations of disease are likely influenced by the areas of the brain that are infected and also by the innate and learned strengths of the child in engaging in social interactions. Less severe congenitally or later acquired childhood illness can explain the increasing prevalence of learning and behavioral problems in school age children and some criminal behaviors in adults.

**Provocation Encephalitis**

Reports of illness occurring shortly after receiving vaccines are officially dismissed on the basis of statistical data that such events may be coincidental. Rarely is serious consideration given to the prospect of vaccine provocation of an underlying stealth adapted virus infection [47]. Purified microbial vaccine antigens are not inherently highly immunogenic and immune-boosting adjuvants are commonly co-administered with the vaccine. The concurrent use of multiple vaccines, each with its own adjuvant, can potentially result in unnecessarily excessive immune stimulation. Stealth adapted viruses may well continue to express components, which in the strong adjuvant setting of vaccination, may become recognizable as antigens by the strengthened cellular immune system. Vaccination could, thereby, provoke an illness, which would likely have neuropsychiatric manifestations. Another scenario can occur with administering live virus vaccines. It is that co-infecting viruses may potentiate the growth and pathogenicity of stealth adapted viruses. It would be worthwhile to analyze the medical history of vaccine damaged children, including recipients of HPV vaccines for evidence of prior illness consistent with a preexisting stealth adapted virus infection. The Public Health system should review the likely excessive use of adjuvants when multiple vaccines are simultaneously administered. It should also examine the benefits of each of the many recommended vaccines in patients, who by history or laboratory testing, show evidence for being stealth adapted virus infected.

**Alternative Cellular Energy (ACE) Pathway**

Probably the most important insight gained from studies on stealth adapted viruses is that the body is not totally dependent upon the immune system to suppress virus infections. Non-immunological suppression of stealth adapted viruses was first noted in virus cultures [48]. The CPE in positive cultures regularly undergoes a repair process, which is reversed by replacing the tissue culture fluid (medium) with fresh medium. The repair is attributed to the accumulation of cell derived materials, which tend to self-assemble into particles, ribbons and threads. Atypical intra- and extracellular structures can also be seen in brain cells from infected humans [17, 45] and inoculated cats [25]. Similar materials are also present on skin and attached to hair of certain patients, presumably forming by the aggregation of materials released in perspiration [49]. The culture-derived and patient derived pigmented materials display various energy related properties. In particular, the particles are; i) electrostatic, consistent with their tendency to self-assemble; ii) fluorescent under ultraviolet (UV) light, especially when interacting with certain dyes, including neutral red and acridine orange; iii) occasionally ferromagnetic and iv) most strikingly capable of preventing reactivation of stealth adapted virus induced CPE when added to the re-feeding medium [48]. A particularly striking observation when examining brain biopsies of infected patients is the very marked disruption of the cells’ mitochondria [16, 45]. Given this, a reasonable assumption is that the materials are allowing cell survival by bypassing the dependency on mitochondrial metabolism. The materials were, accordingly, termed alternative cellular energy (ACE) pigments [48]. An ACE pathway was envisioned as being a source of energy distinct from mitochondria oxidative phosphorylation of food nutrients. Since a few ACE pigment particles could affect the re-feeding medium, it was considered likely that they were mediating their effects by modifying a property of the culture medium.

Gas chromatograph – mass spectroscopy (GC-MS) analyses of ACE pigments showed a diverse range of relatively simple aliphatic and aromatic chemicals. Energy-dispersive x-ray spectroscopy (EDX) indicated that individual particles contain relatively few minerals, but in aggregate many minerals were identified [48]. The cultures were also characterized by lipid production in the form of long needles, linear troughs, crystal, and pyramid shaped structures [42]. The lipid production continued long after all of the cells had died. Similar lipid production has been seen with particles derived from dried perspiration of stealth adapted virus infected patients [48]. The abiotic synthesis of lipids is extremely interesting with regards to the origin of life.

Patient derived ACE particles in water can lead to the slow formation of gas bubbles [49]. Patient and culture derived ACE particles have also been shown to reduce the surface tension and specific heat of water and to increase its volatility (unpublished). These actions have now been explained by the capacity of ACE particles to attract an environmental force termed KELEA (kinetic energy limiting electrostatic attraction) and transfer the energy to water, resulting in a loosening of the intermolecular hydrogen bonding [50]. Once water is sufficiently activated, its separated charges can then directly absorb KELEA from the environment, leading to further activation of the water [51]. As noted above, the ACE pathway is viewed as a naturally occurring mechanism that enables cells to derive energy apart from that provided through food metabolism. The production of readily identifiable ACE pigments is likely the response to an insufficiency of cellular energy (ICE); a cause of which can be the increased energy demands caused by an infection. Evoked
fluorescence within the skin of an elderly patient being treated by neutral red dye for shingles suggested that ACE pigments may be distributed via nerves to localized areas of ICE (unpublished). Transport may also occur within circulating blood cells. The basic notion, however, is that all cells have the potential capacity to utilize the ACE pathway through the production of ACE pigments. The resulting enhanced kinetic activity of the intracellular fluid is thought to facilitate normal metabolic activities and, thereby, help reduce the severity of disease. Conversely, factors that can inhibit the functioning of the ACE pathway may underlie disease exacerbations. An exciting possibility is that electrical activity of the brain may essentially act as a variable antenna, able to absorb KELEA into the body [52]. Indeed, a direct relationship may exist between the brain and activation of the body's fluids. Consuming KELEA activated ACE Water may provide an expedient way to help ensure a positive feedback relationship. These concepts are helping to open a new paradigm of testable therapies, as will be discussed in future articles.

Summary

This review is intended to help lay the foundation for those unfamiliar with the concept of stealth adaptation of viruses. While the existence of stealth adapted viruses poses both political and scientific challenges to Public Health authorities, there is a major role for virologists to help in further characterizing their origins, host range, replication, transmission and pathogenicity. While showing a limitation of the immune system in virus defense, the study of stealth adapted viruses has revealed a novel alternative cellular energy (ACE) pathway that can be utilized in suppressing both stealth adapted viruses and the conventional viruses from which they are derived.

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References

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